

BIOSYNTHESIS OF METELOIDINE FROM 3 α -TIGLOYLOXYTROPANE IN *DATURA INNOXIA*

EDWARD LEETE and DONALD H. LUCAST

Natural Products Laboratory*, School of Chemistry, University of Minnesota,
Minneapolis, MN 55455, U.S.A.

(Received 7 October 1974)

Key Word Index—*Datura innoxia*; Solanaceae; 3 α -tigloyloxytropane; meteloidine; alkaloid biosynthesis.

Abstract—The administration of 3 α -tigloyl-[1- 14 C]-oxytropane-[3 β - 3 H] (3 H/ 14 C = 11.0) to *Datura innoxia* plants for 7 days led to the formation of radioactive meteloidine (3 H/ 14 C = 11.6). Degradation of the meteloidine indicated that the alkaloid was labeled specifically with 3 H at C-3 of its teloidine moiety, and on the carbonyl group of its tigloyl residue with 14 C. These results strongly favor the hypothesis that hydroxylation of tropine occurs after formation of its tigloyl ester.

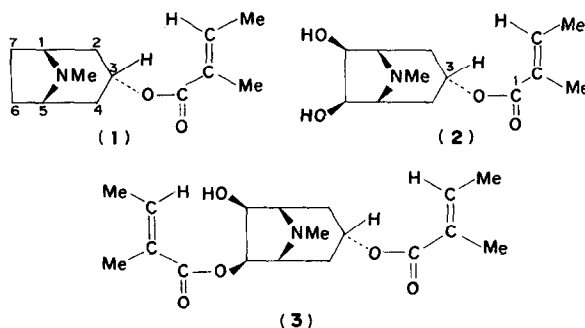
INTRODUCTION

We have previously shown [1] that the teloidine moiety of meteloidine (2) is derived from tropine, hydroxylation occurring at C-6 and C-7. The work of Evans and Woolley [2,3] suggested that this hydroxylation may occur after the formation of the tigloyl ester of tropine. We have now tested this hypothesis by feeding 3 α -tigloyl-[1- 14 C]-oxytropane-[3 β - 3 H] (1) to *Datura innoxia* plants.

RESULTS AND DISCUSSION

Tiglic-[1- 14 C] acid [4] and tropine-[3 β - 3 H] [1] were prepared as previously described. 3 α -Tigloyloxytropane was obtained by heating tigloyl chloride with tropine hydrochloride at 100° [5,6]. 3 α -Tigloyl-[1- 14 C]-oxytropane hydrochloride and 3 α -tigloyloxytropane-[3 β - 3 H] hydrochloride were assayed for radioactivity, mixed and fed to 4-month-old *Datura innoxia* plants by the wick method. The plants were harvested after 7 days. TLC on the crude alkaloids afforded radioactive meteloidine, 7 β -hydroxy-3 α ,6 β -ditigloyloxytropane (3), and 3 α -tigloyloxytropane, activities of these alkaloids being recorded in the Table. The meteloidine was degraded as previously described [1] (see Experimental) and it

was established that essentially all the 3 H was located at the 3 β -position of the teloidine half of the molecule. The tiglic acid contained all the 14 C activity and this was located at the carbonyl group. Since the 3 H/ 14 C ratio in the isolated meteloidine was essentially the same as in the administered 3 α -tigloyloxytropane it seems probable that hydroxylation at C-6 and C-7 occurs on the ester. The recovered 3 α -tigloyloxytropane had low specific activity indicating that it undergoes rapid metabolism in the plant. It contained very little 14 C and this result can be rationalized by postulating a reversible hydrolysis of this ester to tiglic acid and tropine, the latter being then metabolized at a slower rate than tiglic acid.



The incorporation of activity into 7 β -hydroxy-3 α ,6 β -ditigloyloxytropane was lower than that into meteloidine, and the 3 H/ 14 C ratio was much

* Contribution No. 136 from this Laboratory.

Table 1. Activities of the 3 α -Tigloyl-[1-¹⁴C]-oxytropine-[3 β -³H] fed, the isolated alkaloids, and the degradation products of meteloidine

Alkaloids (wt)	Specific activity (dpm/mM)		³ H/ ¹⁴ C	Specific incorporation (³ H, %)
	³ H	¹⁴ C		
Alkaloid fed				
3 α -Tigloyl-[1- ¹⁴ C]-oxytropine-[3 β - ³ H]-HCl (20 mg)	1.34 \times 10 ⁹	1.22 \times 10 ⁸	11.0	
Alkaloids isolated				
Meteloidine (38 mg)	4.75 \times 10 ⁶	4.10 \times 10 ⁵	11.6	0.35
3 α -Tigloyloxytropine (26 mg)	2.27 \times 10 ⁴	<2.0 \times 10 ²	>100	0.0017
7 β -Hydroxy-3 α ,6 β -ditigloyl-oxytropine (18 mg)	1.17 \times 10 ⁶	3.31 \times 10 ⁴	35	0.09
Degradation products of the meteloidine				
Isopropylideneteloidine	4.10 \times 10 ⁶	negligible		
Isopropylideneteloidinone	<0.1 \times 10 ⁶	—		
Tigloyl- α -naphthylamide	6.0 \times 10 ³	3.59 \times 10 ⁵		
Barium carbonate (from the COOH group of tiglic acid)	—	3.3 \times 10 ⁵		

higher than the administered 3 α -tigloyloxytropine. This result suggests that one or both of the tigloyl residues of this alkaloid are labile, reversible hydrolysis and esterification with non-radioactive tiglic acid occurring during the feeding experiment.

EXPERIMENTAL

General methods. A Nuclear Chicago Mark II Liquid scintillation Counter was used for assay of the radioactive compounds using dioxane-EtOH with the usual scintillators [7].

3 α -Tigloyloxytropine-[3 β -³H] hydrochloride. Tigloyl chloride (236 mg, 2 mM) and tropine-[3 β -³H] hydrochloride [1] (190 mg, 3.0 \times 10⁹ dpm, 1.1 mM) were heated on a steam bath for 1.5 hr. Dil. K₂CO₃ (5%) was added to cooled reaction mixture which was then extracted with Et₂O. Residue obtained on evaporation of dried (MgSO₄) extract was treated with a soln of HCl in Et₂O. The resultant solid was crystallized to constant activity from EtOAc affording 3 α -tigloyloxytropine-[3 β -³H] hydrochloride (50 mg), mp 215–216° (lit. [6] mp 214.5–217.5°), having an activity of 2.68 \times 10⁹ dpm/mM.

3 α -Tigloyl-[1-¹⁴C]-oxytropine hydrochloride. Tiglic-[1-¹⁴C]-acid [4] (460 mg, 1.1 \times 10⁹ dpm, 4.6 mM) and SOCl₂ (6 ml) were stirred at room temperature for 30 min and then refluxed for 2 hr. Excess SOCl₂ was removed at red pres and the residual tigloyl chloride heated with tropine hydrochloride (950 mg, 5.53 mM) on a steam bath for 1.5 hr. The 3 α -tigloyl-[1-¹⁴C]-oxytropine hydrochloride was isolated as described in the previous section and had an activity of 2.44 \times 10⁸ dpm/mM.

Administration of 3 α -tigloyl-[1-¹⁴C]-oxytropine-[3 β -³H] hydrochloride to *D. innoxia* and isolation of the alkaloids. The previously described ³H and ¹⁴C labeled 3 α -tigloyloxytropine hydrochloride (10 mg of each) were dissolved in H₂O and fed to 8 4-month-old *D. innoxia* plants growing in soil in a greenhouse, by means of cotton wicks inserted into the stems of plants near to ground level. After 1 week the plants

were harvested (fr. wt 745 g) and macerated in a Waring blender with a mixture of CHCl₃ (1 liter), Et₂O (1 liter), and conc NH₄OH (250 ml). The organic layer was evaporated to 600 ml, and extracted with 2 N H₂SO₄ (6 \times 200 ml). This acid extract was made basic with K₂CO₃ and extracted with CHCl₃. Dried (MgSO₄) extract on evaporation yielded crude alkaloids (215 mg, ³H: 4.63 \times 10⁶ dpm, ¹⁴C: 2.1 \times 10⁵ dpm). Alkaloids were separated by preparative TLC on Si gel PF-254 (Merck) as previously described [1].

Degradation of the meteloidine. The meteloidine isolated from the plant was converted to isopropylideneteloidine and tiglic acid (assayed as tigloyl- α -naphthylamide [8] as previously described [9]). The isopropylideneteloidine was oxidized with CrO₃ in C₅H₅N affording isopropylideneteloidinone. The tiglic acid was subjected to a Schmidt reaction affording CO₂ which was collected and assayed as BaCO₃. The activities of these degradation products are recorded in the Table.

Acknowledgement—This investigation was supported by a research grant GM-13246 from the National Institutes of Health, U.S. Public Health Service.

REFERENCES

1. Leete, E. (1972) *Phytochemistry* **11**, 1713.
2. Evans, W. C. and Woolley, J. G. (1965) *J. Pharm. Pharmacol.* **17**, 375.
3. Woolley, J. G. (1966) *Abhandl. Deut. Akad. Wiss. Berlin, Kl. Chem. Geol. Biol.* (3) 531.
4. Basey, K. and Woolley, J. G. (1973) *Phytochemistry* **12**, 2883.
5. Barger, G., Martin, W. G. and Mitchell, W. (1937) *J. Chem. Soc.* 1820.
6. Leary, J. D., Khanna, K. L., Schwarting, A. E. and Bobbitt, J. M. (1963) *Lloydia* **26**, 44.
7. Friedman, A. R. and Leete, E. (1963) *J. Am. Chem. Soc.* **85**, 2141.
8. Leete, E. (1973) *Phytochemistry* **12**, 2203.
9. Leete, E. and Nelson, S. J. (1969) *Phytochemistry* **8**, 413.